

## THE INFLUENCE OF DIFFERENT LENGTH ON THE CLOROFILINE CONTENT OF *ZEA MAYS* L. PLANTS

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### Abstract

*For plants and some bacteria, light, so electromagnetic wave, is not only a form of energy that drives and facilitates photosynthesis, it is also a sensor of the environment that brings much information but also a stabilizer and regulator of life or different forms of life.*

*Plant development is a process influenced by internal factors such as biological clock and gene expression, as well as external factors such as temperature and light.*

*Light is one of the most important environmental factors in plant growth and growth, a plant with the ability to maximize photosynthesis to evaluate feed-back quality, quantity, and light propagation direction.*

*Light as a decisive factor in plant life and development influences their respiration, perspiration, closure and opening of stomate osteoles, accumulation of chloroplast assimilable pigments, and many other metabolic processes.*

*Higher plants use only visible components of solar energy, those outside the visible spectrum being harmful, solar energy transmitted and received as energy quanta called photons.*

**Key words:** Photosynthetic activity, UV radiations, chlorophyll, light reaction, ATP molecule, chemical reaction, energy.

### INTRODUCTION

Different culture species have the capacity of tolerating UV-B radiation and retaining chlorophyll in leaves, the results varying for monocotyledonous, in comparison to dicotyledonous ones (0-33% in monocotyledonous species, compared to 10-78% in dicotyledonous species). (Tevini et al., 1981)

The variation in the amount of chlorophyll can be attributed to the dosage ratio of UV-B radiation and to the light spectrum (photosynthetic active radiation – PAR), this explaining the degree of damage caused by the UV-B radiation. (Cassi-Lit 1997, Tevini et al., 1991)

The harmful effects on plants caused by the abiotic stress factors in conjunction with the UV stress, is reflected in alterations of the plant's physiology, causing a reduction in their growth and a decrease in their bioproductivity (Khan, 2003).

Chloroplast damage by overexposure to UV-B radiation can lead to the decrease in the chlorophyll content; this involves ultrastructural changes, a decrease of the photosynthetic

In the wild, plants are subjected to certain stress factors, out of which the UV-B radiations (230-320 nm) play an important role, because more and more UV-B reach the Earth's surface due to the depletion of the stratospheric ozone layer. (Deckmyn et al., 1994, Caldwell et al., 1998)

The UV-B radiation causes a net inhibition of the photosynthesis (Tevini 1993), physiological effects including the reduction of carbon assimilation during photosynthesis, the alteration of the stomatal function. (Teramura 1991) Numerous lab studies have shown that this inhibition seems to result from a malfunction in the photosynthetic cycle, it being affected by the gas exchange at the leaf level. (Teramura 1990; Tevini and Teramura et al., 1989)

In our experiments we sought to answer the question whether the treatment with UV-B radiations, of different wavelengths, between 280 – 310 nm, has a stressful effect on plants, resulting in changes in the photosynthetic activity and chlorophyll content, and if there are any differences between the control plants and the treated ones. (Wellmann 1984, Strid, 1994) et al.

All measurements were performed on days 1, 3 and 4 of the treatment, on young plants, the samples consisting of leaves collected from the plants which had the third leaf fully developed. (Ulm R. et al., 2005, Ulm et al., 1994)

Light-dependent stage (light reaction), chlorophyll absorbs light energy, which stimulates some electrons in pigment molecules, transferring them in layers with higher levels of energy. (Marmur, J. 1961, Prewitt, J.M.S. et al., 1970)

They leave the chlorophyll and passed through a series of molecules to form NADPH (enzyme) and the ATP molecule that stores energy. (Sullivan, J.H., et al., 1997)

The oxygen resulting from the chemical reaction is released into the atmosphere through the pores of the leaves. (Prewitt, J.M.S. et al., 1970)

## MATERIAL AND METHOD

Analyzed the effect on photosynthesis through the fluorescent induction, the intracellular carbon dioxide concentration and the relative chlorophyll content measurements.

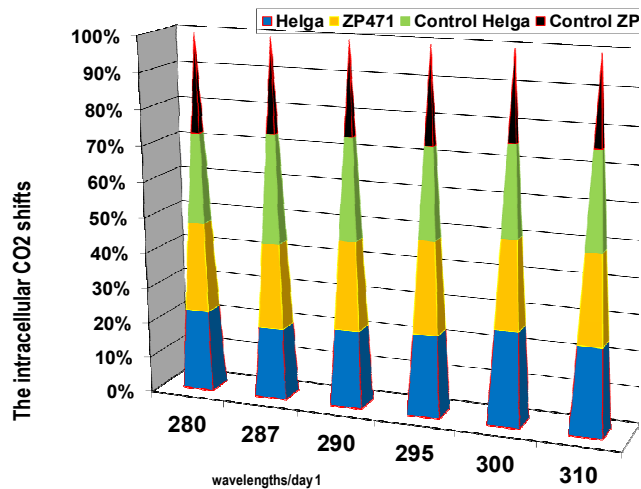
All analyses were performed on days 1, 3 and 4 of the treatment, on young plants, on their third developed leaf.

According to results, it can be stated that, for most low wavelength values, the lowest values of chlorophyll variation and of the CO<sub>2</sub> concentration, of the batch of control plants, have been measured.

## RESULTS AND DISCUSSION

Data from our experiments, showing significant growth, both for the photosynthetic capacity as well as for the intracellular carbon concentration (Fig. 1) assimilated at wavelengths of 287-290 nm confirm the observations from literature, but point out the importance of these readings in selecting tolerant corn genotypes tolerant to UVB, which can be productive in such conditions.

Based on the values in the table we can state that the intracellular CO<sub>2</sub> concentration correlates with the photosynthetic activity for days 3 and 4 of treatment, compared with the control batch, and for days 1 and 3 for the plants treated with the wavelength 287 nm, values were also correlated for both corn hybrids.



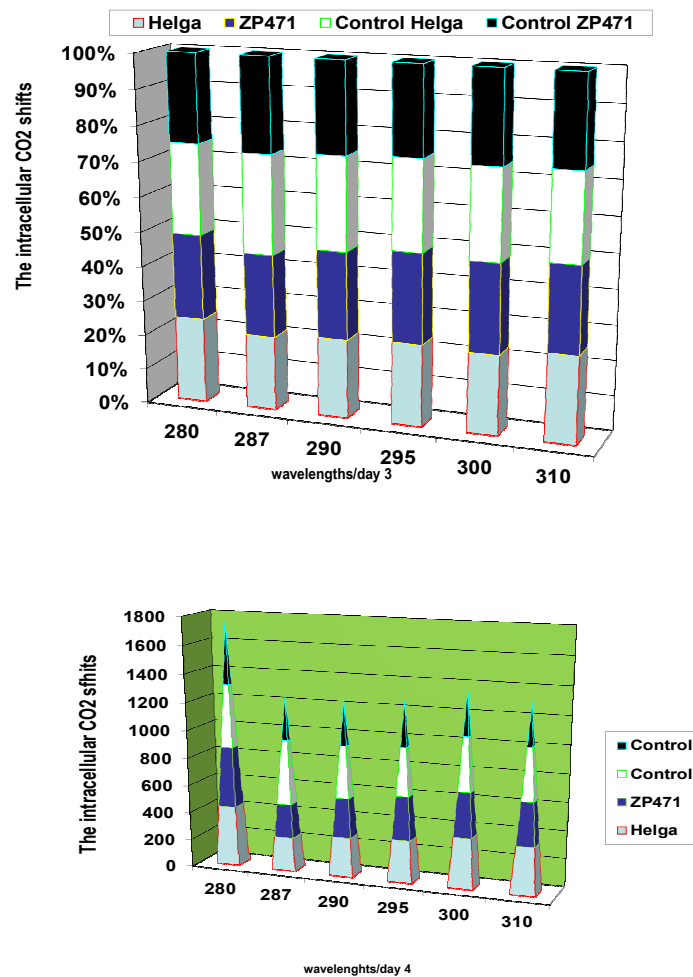


Fig. 1. The intracellular CO<sub>2</sub> shifts (CJ) according to the wavelength changes in day 1 (A), 3(B), 4(C) in corn plants for the control and treated batches. \*significant values in comparison to the control batch at P<0.05

The main protection mechanism against UVB includes the accumulation of compounds that can absorb this radiation. In the case of two wavelengths (285 nm and 287 nm) we have noticed a significant increase of the chlorophyll content in treated plants, in comparison to the treated batch, throughout the 3 days of measurements, after a period of 1 day of wavelengths exposure.

In the case of the two wavelengths (285 nm and 287 nm) there has been noticed a significant increase in the chlorophyll content for the batch of treated plants, in comparison to the control batch, throughout the 3 days of measurements, after the 1 day period of exposure.

After the third day of irradiation with ultraviolet UV-B type radiation, with 280-300 nm wavelengths, a slight change in the chlorophyll content has been noticed, in the case of the corn hybrid ZP471, and equally in the case of the Helga hybrid concerning the control-witness plant, especially at 287 nm.

## CONCLUSIONS

We can conclude that the decrease in the content of chlorophyll ceases after 4 days of irradiation, this being due to the activity of certain antioxidant enzymes, especially the APX enzyme, whose activity increases, thus eliminating the oxidative stress due to wavelengths.

The CO<sub>2</sub> accumulation is linked to the increased photosynthetic capacity.

## REFERENCES

1. Casati, P. and Walbot V., 2004, „Rapid transcriptome responses of maize (Zeamais) to UV-B, in irradiated and shielded tissues.” *Genome* 5, pp. 16
2. Deckmyn G, Martens C., Impens I., 1994, „The importance of the ratio UV-B/photosynthetic active radiation (PAR) during leaf development as determining factor of plant sensitivity to increased UV-B irradiance: effects on growth, gas exchange and pigmentation of bean plants (*Phaseolus vulgaris* cv. Label)”. *Plant Cell Environ* 17: pp. 295-301.
3. Khan W., Prithiviraj B., Smith D., 2003. „Photosynthetic responses of corn and soybean to foliar application of salicylates”. *Journal of Plant Physiology*, 160, pp. 485-492
4. Kramer, P., J., 1983. „Water relations of plants” Academic Press, New York, pp. 34-37
5. Krupa, SV, Kickert, RN. 1989. ”The Greenhouse effect: impacts of ultraviolet-B (UV-B) radiation, carbon dioxide (CO<sub>2</sub>), and ozone (O<sub>3</sub>) on vegetation” *Environ Pollut.* 61(4), pp. 263–393
6. Madronich, S., McKenzie, R.L., Caldwell, M.M., Bjorn, L.O., 1995. „Changes in ultraviolet radiation reaching the earth’s surface”. *Ambio*, pp. 143-152.
7. Mraz C E; Muresan M; Micle O, Vicas L, Pallag A., Coltau M, Puscas I, 2012, Effect of vitamin D on carbonic anhydrase activity experimental research in vitro and in vivo, *Farmacia*,; **60**(2): pp. 264-271.
8. Marmur, J. 1961. „A procedure for the isolation of deoxyribonucleic acid from microorganisms”. *J.Mol. Biol.* 3, pp.208-218.
9. McCloskey JT, Oris JT. 1993. „Effect of anthracene and solar ultraviolet radiation exposure on gill ATPase and selected hematologic measurements in the bluegill sunfish (*Lepomis macrochirus*)” *Aquat Toxicol.* pp. 207–218
10. Prakken, R., 1959. „Induced mutation”. *Euphytica* 8: pp. 270-322.
11. Prewitt, J.M.S. 1970. „Object enhancement and extraction” In A., Rosenfeld and B.S. Lipkin, editors, *Picture Processing and Psychophysics*, Academic Press, New York, pp. 75-149,
12. Pallag A, Jurca T, Sirbu V, Honiges A, Jurca C, 2018 Analysis of the Amount of Polyphenols, Flavonoids and Assessment of the Antioxidant Capacity of Frozen Fruits, *REV CHIM. (Bucharest)*,; **69**(2): pp. 445-448.

13. Pallag A, Jurca T., Pasca B., Sirbu V., Honiges A., Costuleanu M., 2016 Analysis of Phenolic Compounds Composition by HPLC and Assessment of Antioxidant Capacity in *Equisetum arvense* L. Extracts REV CHIM. (Bucharest), , **67**(8): pp 1623-1627
14. Pallag A., Pașca B., Jurca T., Suciuc R., Nemeth S., Vicaș L., 2016 Comparative histo-anatomical researches on the vegetative organs and assessment of antioxidant capacity of two species from *Equisetum* Genus. Farmacia,; **64**(3): pp. 372-377.
15. Pallag A , Gabriela Adriana Filip , Diana Olteanu , Simona Clichici, Ioana Baldea, Tunde Jurca, Otilia Micle, Laura Vicaș , Eleonora Marian , Olga Sorișău , Mihai Cenariu , Mariana Mureșan, 2018, *Equisetum arvense* L. Extract Induces Antibacterial Activity and Modulates Oxidative Stress, Inflammation, and Apoptosis in Endothelial Vascular Cells Exposed to Hyperosmotic Stress, Hindawi, Oxidative Medicine and Cellular Longevity, , Article ID 3060525, pp14
16. Rout, G.R., Jain, S.M., 2004, Micropropagation of ornamental plants—cut flowers. In: Propagation of Ornamental Plants, 4 (2), pp. 3 – 28
17. Street, H.E., 1977, Laboratory organization. In: HE Street, Ed, Plant Tissue and Cell Culture. Univ California Press, Berkeley, Los Angeles, pp. 11 - 30.
18. Strid, Å., Chow, W., Anderson, J. M., 1994, „UV-B damage and protection at the molecular level in plants”. Photosynthesis Research 39: pp.475-489
19. Sullivan, J.H., 1997. „Effects of increasing UV-B radiation and atmospheric CO<sub>2</sub> on photosynthesis and growth: implications for terrestrial ecosystems.” Plant Ecol. 128, pp. 194–206.
20. Sullivan, J.H., Rozema, J., 1999. „UV-B effects on terrestrial plant growth and photosynthesis”. In: Rozema, J. (Ed.), Stratospheric Ozone Depletion: The Effects of Enhanced UV-B Radiation on Terrestrial Ecosystems. Backhuys Publishers, Leiden, pp. 39–57
21. Teramura, A. H., Sullivan, J.H., Ziska, L.H., 1990., „Interaction of elevated ultraviolet-B radiation and CO<sub>2</sub> on productivity and photosynthetic characteristic in wheat, rice, and soybean”. Plant Physiology 94, pp. 470-475
22. Tevini, M., Teramura, A. H., 1989. “UV-B effects on terrestrial plants” Photochemistry Photobiology 50, pp. 479-487.
23. Tosserams, M., 1998, „Stratospheric ozone depletion – responses of dune grassland plants and faba bean to ultraviolet-B radiation”. Drukwerk: Ponsen & Looijen BV, Wageningen, pp.3-11.
24. Ulm, R., Nagy, F. 2005. „Signalling and gene regulation in response to ultraviolet light.” Curr. Opin. Plant Biol. 8, pp. 477-482.
25. Webb, Ann R., 1991, „Solar UV-B radiation measurement at the earth’s surface: techniques and trends. Impact of global climatic changes on Photosynthesis and Plant Productivity”, Oxford & IBH Publishing CO.PVT.LTD, pp.23-34.
26. Wellmann, E., Beggs, C.J., Möhle, B., Schneider-Ziebert, U., Steinmeitz, V., 1984. „Plant responses to solar UV-B radiation”. Proc. 2nd European Symposium on Life Sciences Research in Space, Germany, pp. 61-65.
27. Quate, F.E., Takayanagi, S., Ruffini, J., Sutherland, J.C., and Sutherland, B.M., 1994. „DNA damage levels determine cyclobutyl pyrimidine dimer repair mechanisms in alfalfa seedlings”. Plant Cell 6, pp. 163-164.